



## Mechanistic study of endothelium independent vasodilation effects of wogonin

Qi Wu<sup>1#</sup>, Wanlong Zhao<sup>2#</sup>, Gangling Chen<sup>3\*</sup>, Anran Yao<sup>3</sup>, Cheng Yang<sup>3</sup>, Yue Zhao<sup>3</sup> & Jiangwei Zhang<sup>3</sup>

<sup>1</sup>State Key Laboratory of Natural Medicines, Research Department of Pharmacognosy, School of Traditional Chinese Pharmacy, China Pharmaceutical University, Nanjing 211198, China

<sup>2</sup>Shanghai Qingpu Institute for Food and Drug Control, Shanghai 201799, China

<sup>3</sup>Department of Pharmacology of Chinese Materia Medica, School of Traditional Chinese Pharmacy, China Pharmaceutical University, Nanjing 211198, China

Received 22 October 2020; revised 31 October 2021

*Scutellaria baicalensis* Georgi, locally known as HuangQin, and commonly as Baikal or Chinese skullcap, is an important herb in Chinese traditional medicine. The flavonoids from this plant are main active substances responsible for its medicinal applications. Wogonin is one such active ingredient derived from this plant. Here, we investigated the mechanism of the vasodilation effect of wogonin on isolated rat thoracic aortas. For this study, endothelium intact and endothelium removed thoracic aortic rings were prepared from rats. Using a tension transducer, the tension of the rat thoracic aortic rings was recorded. Results showed that wogonin is able to relax the endothelium-intact aortic rings, but L-NAME, indomethacin (Indo), and methylene blue (MB) could not reduce the tension in these rings. Wogonin was also able to relax endothelium-removed rings. However, treatment with tetraethylammonium (TEA), BaCl<sub>2</sub>, glibenclamide (Gly), 4-aminopyridine (4-AP), and verapamil (Ver) had no effect on vasodilation induced by wogonin. Using wogonin to pre-treat endothelium-removed aortic rings reduced the contraction induced by K<sup>+</sup>. Pre-treatment of endothelium-removed aortic rings with wogonin markedly reduced the contraction induced by 10<sup>-6</sup> M PE in Ca<sup>2+</sup>-free solution. It could be concluded that L-type calcium channels and intracellular Ca<sup>2+</sup> release is inhibited by wogonin.

**Keywords:** Baikal skullcap, Chinese skullcap, Herbal, Intracellular calcium release, Ca<sup>2+</sup> channel

Cardiovascular Disease (CVD) is heart and circulatory system disorders, which has become a major human health issue today<sup>1,2</sup>. Medicines with vasodilatory effects are useful in preventing and curing this type of disease<sup>3-5</sup>. Several natural medicines have been reported that could potentially prevent CVDs<sup>6,7</sup>. *Scutellaria baicalensis* Georgi (Lamiaceae), a Chinese medicinal plant commonly called Baikal skullcap or Chinese skullcap, has heat dampness, purging fire detoxification, and tocolysis characteristics and can be used to staunch bleeding based on the theory of traditional Chinese medicine<sup>8,9</sup>. Pharmacological studies have shown that it protects endothelial cells<sup>10</sup> and exhibit anticancer<sup>11,12</sup>, antivirus<sup>13</sup>, anti-inflammatory effects<sup>14,15</sup>, and has potential application in the treatment of CVDs<sup>16,17</sup> in addition to hepatitis, diarrhoea, vomiting and high blood pressure<sup>8</sup>. There are several flavone derivatives reported from the root of *S. baicalensis*, such as wogonin, wogonoside, baicalein and baicalin. Among those flavone derivatives, wogonin (C<sub>16</sub>H<sub>12</sub>O<sub>5</sub>) has

attracted the attention of lots of researchers (Scheme 1). It is reported that wogonin has therapeutic effects on ischemic brain injury<sup>18</sup>, vascular inflammatory<sup>19</sup>, antitumor *in vivo* and *in vitro*<sup>20,21</sup>, in particular antithrombotic activities<sup>22</sup> and anti-angiogenesis<sup>23</sup>. Qu *et al.*<sup>24</sup> have reported that wogonin can exert vasodilatory effect on isolated rat aortic rings by endothelium-independent pathway, while the exact mechanisms still remain unclear.

Based on these pharmacological effects, in this study, we have investigated whether wogonin can affect vascular ion channels or factors. We used isolated rat aortic rings to test the effect of wogonin on vasodilation.

### Materials and Methods

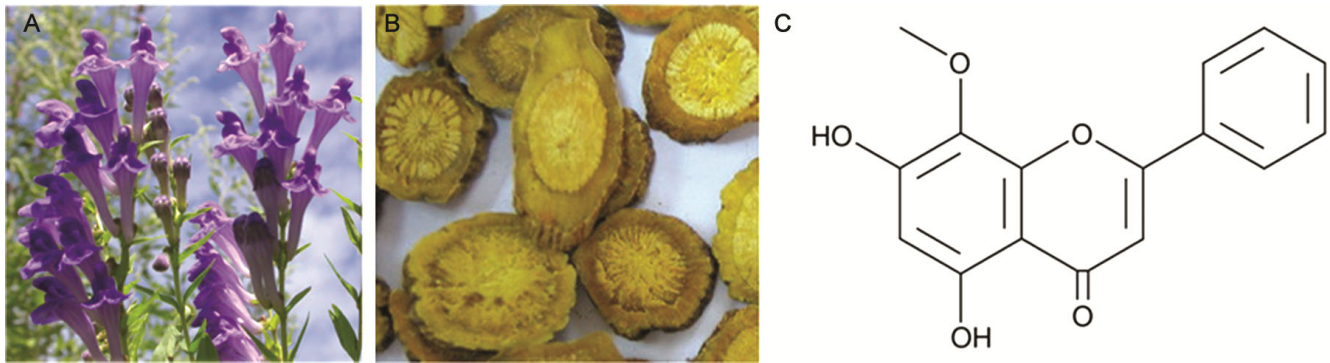
#### Experimental drug

Acetylcholine (ACh), L-NG-nitroarginine methyl ester (L-NAME), indomethacin (Indo), Glibenclamide (Gly) and Tetraethylammonium (TEA, >98% purity) were purchased from Sigma Aldrich (St. Louis, MO, USA). Phenylephrine hydrochloride (PE, >98% purity), 4-aminopyridine (4-AP, >99% purity) was purchased from TCI Development Co., Ltd

\*Correspondence:

E-Mail: chengangling@cpu.edu.cn

#Equal contributed



Scheme 1 — (A) *Scutellaria baicalensis* Georgi; (B) decoction pieces of *S. baicalensis*; and (C) Chemical structural formula of wogonin.

(Shanghai, China). Wogonin ( $C_{16}H_{12}O_5$ , MW 284.26, 99.01% purity) was bought from the Chengdu Pusi Bio-Technology Co., Ltd (Chengdu, Sichuan, China) Ethylenediaminetetraacetic acid tetrasodium salt (EDTA·4Na, 99-102% pure), verapamil (Ver, 99% pure) and methylene blue (MB, 99% pure) were obtained from Aladdin Industrial Corporation (Shanghai, China). All other reagents used were of analytical purity. ACh and PE were dissolved in deionized water, whereas wogonin was dissolved in DMSO. The Krebs-Henseleit (K-H) solution consisted of  $118 \text{ mmol}\cdot\text{L}^{-1}$  NaCl,  $25 \text{ mmol}\cdot\text{L}^{-1}$   $\text{NaHCO}_3$ ,  $4.7 \text{ mmol}\cdot\text{L}^{-1}$  KCl,  $2.5 \text{ mmol}\cdot\text{L}^{-1}$   $\text{CaCl}_2$ ,  $1.2 \text{ mmol}\cdot\text{L}^{-1}$   $\text{MgCl}_2$ ,  $1.2 \text{ mmol}\cdot\text{L}^{-1}$   $\text{KH}_2\text{PO}_4$ , and  $2.2 \text{ mmol}\cdot\text{L}^{-1}$  glucose (pH = 7.4). The  $\text{Ca}^{2+}$ -free K-H solution and  $\text{Ca}^{2+}$ -free high concentration  $\text{K}^+$  K-H solution ( $60 \text{ mmol}\cdot\text{L}^{-1}$   $\text{K}^+$ ) both containing  $1\times 10^{-4} \text{ mol}\cdot\text{L}^{-1}$  EDTA.

#### Animals and aortic ring preparation

Male Sprague-Dawley rats weighing 200-230 g were provided by the Comparative Medicine Centre of Yangzhou University (Yangzhou, China). Animals were housed in a temperature and light controlled room ( $23\pm 1^\circ\text{C}$ ; 12 h light/dark cycle) and allowed to freely access food and water. All animal experiments were strictly conducted following a protocol approved by the Animal Ethics Committee of China Pharmaceutical University.

Rats were euthanized by cervical dislocation under diethyl ether anesthesia. To isolate aortic ring, the thoracic aortas were removed immediately and immersed in K-H solution, then the attached tissue were removed. The thoracic aortas were sheared into aortic rings (3 mm long). A wooden toothpick was used to gently rub the inner portion of the aortic rings to displace the endothelial layer. The endothelium was deemed intact if  $10^{-5} \text{ mol}\cdot\text{L}^{-1}$  of ACh induced

80% relaxation, which was pre-contracted by  $10^{-6} \text{ mol}\cdot\text{L}^{-1}$  of PE. A relaxation expressed  $<5\%$  was recorded when the endothelium was completely denuded.

The aortic rings were suspended in the K-H solution bath (5 mL), which maintained with 95%  $\text{O}_2$  and 5%  $\text{CO}_2$  at  $37^\circ\text{C}$ . One side of the ring was connected to an L-shape hook, and the other side was connected to the tension transducer connected to a MedLab BL-420 Polygraph (Tai Meng Technology, Chengdu, China). The baseline tension loaded onto the aortic rings was 2 g. Each ring was equilibrated in the bath solution for 30 min before involving the contractile response to the addition of  $60 \text{ mmol}\cdot\text{L}^{-1}$  of the high  $\text{K}^+$  K-H solution. The aortic rings were washed thrice with the K-H solution, each time for 10 min. Recorded the relaxant response of the ACh concentration gradient ( $10^{-9}$ ,  $10^{-8}$ ,  $10^{-7}$ ,  $10^{-6}$  and  $10^{-5} \text{ mol}\cdot\text{L}^{-1}$  ACh) after pre-contraction was induced by  $10^{-6} \text{ mol}\cdot\text{L}^{-1}$  of PE. Relaxation was expressed as Relaxation %. Relaxation % = [(Maximal tension of  $10^{-6} \text{ mol}\cdot\text{L}^{-1}$  PE - Minimal tension of X)/(Maximal tension of  $10^{-6} \text{ mol}\cdot\text{L}^{-1}$  PE - resting tension)]\*100%, Minimal tension of X is the minimum tension relaxed by reagent.

#### Vasodilation induced by wogonin and the effect of wogonin on endothelium function

To test the effect of wogonin on endothelium intact aortic rings, increasing concentrations of wogonin were applied to the rings after pre-contracting with  $10^{-6} \text{ mol}\cdot\text{L}^{-1}$  of PE. L-NAME ( $10^{-4} \text{ mol}\cdot\text{L}^{-1}$ ), Indo ( $10^{-5} \text{ mol}\cdot\text{L}^{-1}$ ), and MB ( $10^{-5} \text{ mol}\cdot\text{L}^{-1}$ ) were used to test the mechanism of the endothelium relaxation pathways.

#### Vasodilation induced by wogonin and the role of $\text{K}^+$ channels

To determine whether  $\text{K}^+$  channels play a role in the vasodilation effect of wogonin, endothelium-removed

aortic rings were pre-incubated with TEA (calcium-activated  $K^+$  channel blocker,  $3 \times 10^{-3} \text{ mol} \cdot \text{L}^{-1}$ ), Gly (ATP-sensitive  $K^+$  channel blocker,  $10^{-5} \text{ mol} \cdot \text{L}^{-1}$ ), 4-AP (voltage-dependent  $K^+$  channel blocker,  $10^{-4} \text{ mol} \cdot \text{L}^{-1}$ ), or  $\text{BaCl}_2$  (inward rectifier of  $K^+$  channels, KIR,  $10^{-4} \text{ mol} \cdot \text{L}^{-1}$ ) for 20 min. Wogonin (1, 3, 10, 30, 100 and 300  $\mu\text{mol} \cdot \text{L}^{-1}$ ) was added cumulatively after PE ( $10^{-6} \text{ mol} \cdot \text{L}^{-1}$ ) was addition,

#### Vasodilation induced by wogonin and the role of $\text{Ca}^{2+}$ channels

To illustrate the relationship between  $\text{Ca}^{2+}$  channels and the pharmacological effects of wogonin, endothelium removed aortic rings were pre-incubated with Ver ( $10^{-5} \text{ mol} \cdot \text{L}^{-1}$ ) for 20 min.  $10^{-6} \text{ mol} \cdot \text{L}^{-1}$  of PE was then added to the bath to induce vasoconstriction, and wogonin (1, 3, 10, 30, 100 and 300  $\mu\text{mol} \cdot \text{L}^{-1}$ ) was added cumulatively.

#### The effect of wogonin on PE- or KCl-induced vasoconstriction in endothelium-removed rings

After equilibrated in K-H solution and pre-incubated with wogonin ( $28.58 \mu\text{mol} \cdot \text{L}^{-1}$  or  $85.74 \mu\text{mol} \cdot \text{L}^{-1}$ ) for 10 min, endothelium-removed rings were exposed to PE ( $10^{-9}$ ,  $10^{-8}$ ,  $10^{-7}$ ,  $10^{-6}$ , or  $10^{-5} \text{ mol} \cdot \text{L}^{-1}$ ) or KCl ( $1 \times 10^{-2}$ ,  $2 \times 10^{-2}$ ,  $4 \times 10^{-2}$  or  $8 \times 10^{-2} \text{ mol} \cdot \text{L}^{-1}$ ). PE- or KCl induced vasoconstriction were expressed as contraction%. For PE, Contraction % = (Maximal tension of PE – resting tension)/(Maximal tension of 60 mM  $K^+$  – resting tension)\*100%. For KCl, Contraction % = (Maximal tension of KCl – resting tension)/(Maximal tension of  $8 \times 10^{-2} \text{ mol} \cdot \text{L}^{-1} K^+$  – resting tension)\*100%.

#### The effect of wogonin on intracellular calcium release

To evaluate the effect of wogonin on intracellular calcium release during vasoconstriction, endothelium-removed rings were washed thrice with  $\text{Ca}^{2+}$ -free K-H solution and incubated for 10 min, followed by addition of  $10^{-6} \text{ mol} \cdot \text{L}^{-1}$  of PE. The rings were then washed thrice with K-H solution and equilibrated for 40 min to refill the intracellular  $\text{Ca}^{2+}$  stores. Finally, the rings were washed with  $\text{Ca}^{2+}$ -free K-H solution thrice and pre-incubated with 28.58 or 85.74  $\mu\text{mol} \cdot \text{L}^{-1}$  of wogonin for 10 min before  $10^{-6} \text{ mol} \cdot \text{L}^{-1}$  of PE was added to induce the second contraction.

#### Data analysis

All data are expressed as the mean  $\pm$  SD. Statistical significance was evaluated using an ANOVA followed by Dunnett's test.  $P < 0.05$  was taken as a significant difference.  $\text{EC}_{50}$  values were determined by Graphpad Prism 5.

## Results

#### Role of the endothelium in the function of wogonin

Wogonin (1, 3, 10, 30, 100 and 300  $\mu\text{mol} \cdot \text{L}^{-1}$ ) reduced the contraction of endothelium intact aortic rings caused by  $10^{-6} \text{ mol} \cdot \text{L}^{-1}$  of phenylephrine (PE). Pre-treatment of aortic rings with L-NAME, Indo, or MB had no effect on the relaxation induced by wogonin (Fig. 1). L-NAME is a nonselective inhibitor of nitric oxide synthase, while Indo can inhibit cyclooxygenase activity by reducing the generation of prostacyclin. MB is a guanylate cyclase inhibitor. These results indicate that the vasodilatory effect of wogonin is independent of endothelium.

#### Role of $K^+$ channels in the function of wogonin

Wogonin reduced the tension in endothelium-removed aortic rings. The  $\text{EC}_{50}$  values of wogonin were calculated as the compound concentration inducing 50% of the maximum ( $\text{EC}_{50} = 28.58 \mu\text{mol} \cdot \text{L}^{-1}$ ). Endothelium-removed aortic rings were then treated with 4-AP, TEA, Gly and  $\text{BaCl}_2$ . These four blockers had no effect on the function of wogonin (Fig. 2A).

#### Role of $\text{Ca}^{2+}$ channels in the vasodilation effect of wogonin

Contraction of endothelium-removed aortic rings was induced by the addition of  $10^{-6} \text{ mol} \cdot \text{L}^{-1}$  of PE, but addition of Ver was unable to influence vasodilation induced by 1, 3, 10, 30 or 100  $\mu\text{mol} \cdot \text{L}^{-1}$  of wogonin (Fig. 2B). This result indicates that Cav1.2 does not participate in relaxation induced by wogonin. In this study, the function of the cell surface, voltage-gated  $\text{Ca}^{2+}$  channels (VGCCs) were also studied. An

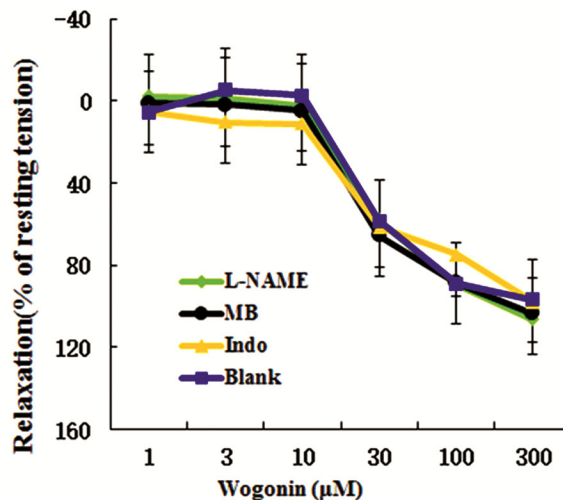


Fig. 1 — Vasorelaxation effects of wogonin on intact endothelium. [Thoracic aortic rings were pre-incubated with L-NAME, Indo, or MB before vasoconstriction induced by PE. Wogonin was then added to induce vasodilation. Data are expressed as the mean  $\pm$  SD,  $n=4-6$ ]

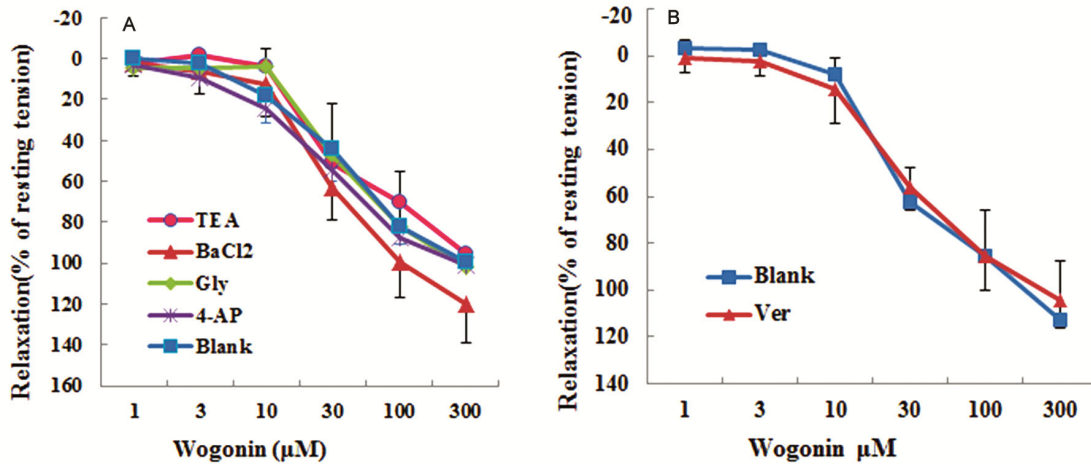


Fig. 2 — Influence of (A) potassium; and (B) calcium channel blockers on the effects of wogonin in vessels lacking endothelium. Thoracic aortic rings were pre-incubated with (A) TEA, BaCl<sub>2</sub>, Gly, or 4-AP; and (B) Ver before vasoconstriction induced by PE. Wogonin was then added to induce vasodilation. [Data are expressed as the mean  $\pm$  SD, n=4-6]

experiment was conducted to investigate the effect of wogonin on the contractions induced by increasing concentrations of KCl. The results showed that 28.58 or 85.74  $\mu\text{mol}\cdot\text{L}^{-1}$  of wogonin reduced contractions induced by KCl (0.01, 0.02, 0.04 or 0.08  $\text{mol}\cdot\text{L}^{-1}$ ) (Fig. 3). The contractions induced by high concentrations of K<sup>+</sup> rely on VGCCs and the opening of Ca<sup>2+</sup> release channels (ryanodine and IP<sub>3</sub> receptors). Thus, wogonin has an effect on the VGCCs or the opening of Ca<sup>2+</sup> release channels.

#### Receptor-mediated Ca<sup>2+</sup> influx

In endothelium-removed aortic rings, wogonin (28.58 or 85.74  $\mu\text{mol}\cdot\text{L}^{-1}$ ) reduced the contraction caused by increasing concentrations of PE (Fig. 4). The contraction induced by PE is mainly associated with receptor-mediated Ca<sup>2+</sup> influx. It is suggested that wogonin could affect Ca<sup>2+</sup> influx.

#### Wogonin affects intracellular calcium release

The endothelium-removed aortic rings were used to test the effects of wogonin on intracellular calcium release. First, wogonin (28.58 or 85.74  $\mu\text{mol}\cdot\text{L}^{-1}$ ) was incubated with endothelium-removed aortic rings in calcium-free K-H solution for 10 min. Next, 10<sup>-6</sup>  $\text{mol}\cdot\text{L}^{-1}$  of PE was added to induce contraction. Both concentrations of wogonin were able to reduce the PE-induced contraction, which was caused by intracellular calcium release (Fig. 5). These results indicate that wogonin inhibits intracellular calcium release.

## Discussion

### Role of the endothelium in wogonin-mediated vasodilation of aortic rings

For endothelium intact aorta, the endothelium, and in particular the endothelial cells, are important

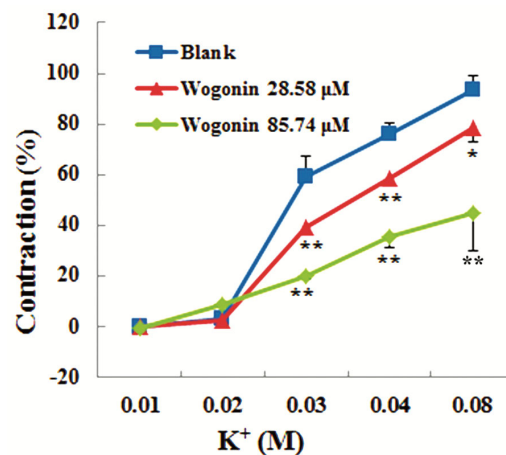


Fig. 3 — Effects of wogonin on vasodilation curves of PE in vessels lacking an endothelium. [Wogonin (28.58 or 85.74  $\mu\text{mol}\cdot\text{L}^{-1}$ ) was added to pre-treat the rings, and then increasing concentrations of PE were added to induce vasoconstriction. Data are expressed as the mean  $\pm$  SD, n=4-6; \*\**P* < 0.01 vs. Blank]

components of the vasodilation process. Endothelial cells release several factors, such as nitric oxide (NO) and prostacyclin (PGI<sub>2</sub>) to relax the aorta<sup>25-28</sup>. NO released by endothelial cells activates guanylyl cyclase and elevates intracellular cyclic guanosine monophosphate (cGMP) synthesis from Guanosine triphosphate (GTP) to exert its biological effects<sup>29,30</sup>. In this study, wogonin relaxed the endothelium-intact aortic rings, which were contracted by prior addition of PE (10<sup>-6</sup>  $\text{mol}\cdot\text{L}^{-1}$ ). This effect could not be blocked by L-NAME, an inhibitor of endothelial nitric oxide synthase (eNOS), indicating that wogonin relax the aorta via an NO independent pathway. Similarly, Indo had no effect on wogonin mediated vasodilation. Indo

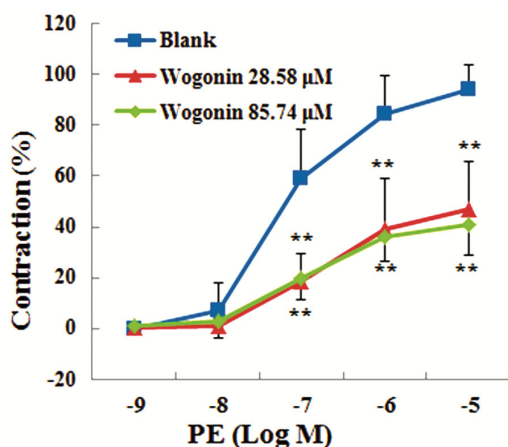


Fig. 4 — Effects of wogonin on the vasoconstriction curves of KCl in vessels lacking an endothelium. [Wogonin (28.58 or 85.74  $\mu\text{mol}\cdot\text{L}^{-1}$ ) was added to pre-treat the aortic rings, and then increasing concentrations of KCl were added to induce vasoconstriction. Data are expressed as the mean  $\pm$  SD, n=4-6; \* $P$  < 0.05, \*\*  $P$  < 0.01 vs. Blank]

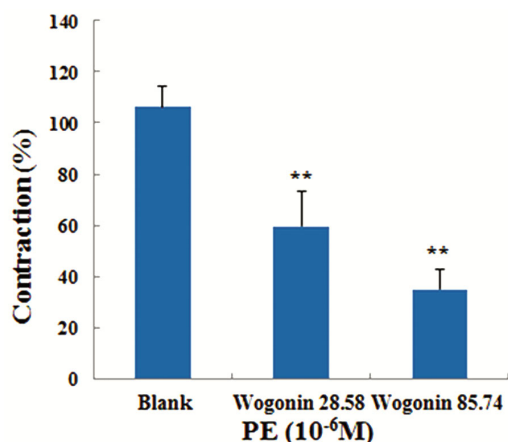


Fig. 5 — Effects of wogonin on intracellular  $\text{Ca}^{2+}$  in aortic rings lacking an endothelium. [Aortic rings were pre-treated with wogonin (28.58 or 85.74  $\mu\text{mol}\cdot\text{L}^{-1}$ ) in  $\text{Ca}^{2+}$ -free K-H solution. PE was then added to induce vasoconstriction. Data are expressed as the mean  $\pm$  SD, n=4-6; \*\*  $P$  < 0.01 vs. Blank]

inhibits cyclooxygenase to reduce the production of  $\text{PGI}_2$ . Thus, the vasodilatory effects of wogonin is independent of prostacyclin ( $\text{PGI}_2$ ). To further verify the results described above, we used methylene blue (MB), a guanylate cyclase inhibitor, to test the role of guanylate cyclase in the vasodilation effect of wogonin. Results showed that MB can not affect the vasodilatory effect of wogonin.

#### Function of smooth muscle pathways in the mechanism of wogonin

$\text{K}^+$  channels,  $\text{Ca}^{2+}$  channels and intracellular calcium release are all vital for vasoconstriction and

vasodilation of endothelium-removed aortic rings<sup>31,32</sup>. In the vascular smooth muscle, there are 4 types of  $\text{K}^+$  channels: voltage dependent  $\text{K}^+$  channels<sup>33</sup>, ATP sensitive  $\text{K}^+$  channels<sup>34</sup>, calcium activated  $\text{K}^+$  channels<sup>35</sup>, and inward rectifier potassium channels<sup>36</sup>. In this study, we used 4-AP, TEA, Gly and  $\text{BaCl}_2$  to block the all four types of  $\text{K}^+$  channels, respectively. The results showed that these four blockers had no effect on the function of wogonin. It can be concluded that the vasodilation function of wogonin is independent of these four types of  $\text{K}^+$  channels.

In this study, verapamil, a Cav1.2 blocker<sup>37,38</sup>, particularly the voltage dependent, L-type, alpha1C subunit (Cav1.2), was used to test the effect of wogonin on endothelium removed aortic rings. The results showed that Ver had no effect on the vasodilation induced by wogonin when rings were pre-contracted with PE ( $10^{-6} \text{ mol}\cdot\text{L}^{-1}$ ). Clearly, wogonin mediated vasodilation is independent with Cav1.2. When pre-incubated endothelium-removed aortic rings with wogonin (28.58 or 85.74  $\mu\text{mol}\cdot\text{L}^{-1}$  for 20 min), the vasoconstriction caused by increasing concentrations of KCl and PE was reduced. Vasoconstriction caused by high concentrations  $\text{K}^+$  is mediated by VGCCs and the opening of  $\text{Ca}^{2+}$  release channels (ryanodine and IP3 receptors)<sup>39,40</sup>. The vasoconstriction caused by PE is mainly a result of receptor-mediated  $\text{Ca}^{2+}$  influx. Together, these results indicate that wogonin exerts its vasodilation effects through the opening of  $\text{Ca}^{2+}$  release channels, as well as the receptor operated  $\text{Ca}^{2+}$  influxes. The blocking effect of low dose of wogonin (28.58  $\mu\text{M}$ ) on the opening of  $\text{Ca}^{2+}$  release channels is slightly weaker than the receptor-mediated  $\text{Ca}^{2+}$  influx. When the dose of wogonin is increased to 85.74  $\mu\text{M}$ , the blocking effect increased and the effect on the two kinds of channels was similar. The effects of wogonin on intracellular calcium release was tested by adding PE ( $10^{-6} \text{ mol}\cdot\text{L}^{-1}$ )<sup>41</sup>, which causes contraction in calcium free K-H solutions. Results showed that wogonin reduced PE-mediated contraction, suggesting that wogonin could block intracellular calcium release. Wogonin 28.58  $\mu\text{M}$  and 85.74  $\mu\text{M}$  both could block the release of intracellular calcium, and the effect of 85.74  $\mu\text{M}$  is stronger. The vasodilatory effect of wogonin on isolated rat aortic rings and the rat uterine smooth muscle also confirmed by other researcher's study<sup>24,42</sup>, but more information of wogonin was studied in this research work, summarized the following points: Confirmed

the effect of wogonin to prostacyclin, guanylate cyclase, the 4 types of K<sup>+</sup> channels, Cav1.2 and the receptor-mediated Ca<sup>2+</sup> influx.

### Conclusion

Based on the results, it could be concluded that wogonin exerts vasodilation effect via blocking the opening of Ca<sup>2+</sup> release channels, receptor operated Ca<sup>2+</sup> influxes and intracellular calcium release. The present findings will be meaningful for revealing the pharmacological effects of wogonin on the cardiovascular system.

### Acknowledgement

This work was mainly supported by the National Natural Science Foundation of China (No. 81503284), the "Double First-Class" University project (CPU2018GF06, CPU2018GF07), Fundamental Research Funds for the Central Universities (No. 2015PY016), and Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD).

### Conflict of Interest

Authors declare no competing interests.

### References

- Virani SS, Alonso A, Benjamin EJ, Bittencourt MS, Callaway CW, Carson AP, Chamberlain AM, Chang AR, Cheng S, Delling FN, Djousse L, Elkind MSV, Ferguson JF, Fornage M, Khan SS, Kissela BM, Knutson KL, Kwan TW, Lackland DT, Lewis TT, Lichtman JH, Longenecker CT, Loop MS, Lutsey PL, Martin SS, Matsushita K, Moran AE, Mussolino ME, Perak AM, Rosamond WD, Roth GA, Sampson UKA, Satou GM, Schroeder EB, Shah SH, Shay CM, Spartano NL, Stokes A, Tirschwell DL, VanWagner LB & Tsao CW, Heart Disease and Stroke Statistics-2020 Update: A Report From the American Heart Association. *Circulation*, 141 (2020) e139.
- Chen G, Thakkar M, Robinson C & Doré S, Limb Remote Ischemic Conditioning: Mechanisms, Anesthetics, and the Potential for Expanding Therapeutic Options. *Front Neurol*, 9 (2018) 40.
- Tetty CO, Yang IJ & Shin HM, Vasodilatory effect of kaempferol-7-O- $\alpha$ -L-rhamnopyranoside via NO-cGMP-PKG signaling. *Arch Biochem Biophys*, 667 (2019) 1.
- Leurgans TM, Bloksgaard M, Irmukhamedov A, Riber LP & De Mey J, Relaxing Responses to Hydrogen Peroxide and Nitric Oxide in Human Pericardial Resistance Arteries Stimulated with Endothelin-1. *Basic Clin Pharmacol Toxicol*, 122 (2018) 74.
- Ferreira-Filho ES, Arcanjo DD, Moura LH, da Silva-Filho JC, Paulino ET, Ribeiro EA, Chaves MH, Oliveira Rde C & de Oliveira AP, Antihypertensive and vasorelaxant effects of ethanol extract of stem barks from *Zanthoxylum rhoifolium* Lam. in rats. *Indian J Exp Biol*, 51 (2013) 661.
- Irfan M, Kim M & Rhee MH, Anti-platelet role of Korean ginseng and ginsenosides in cardiovascular diseases. *J Ginseng Res*, 44 (2020) 24.
- Zhou K, Chen J, Wu J, Wu Q, Jia C, Xu YXZ, Chen L, Tu W, Yang G, Kong J, Kou J & Jiang S, Atractylenolide III ameliorates cerebral ischemic injury and neuroinflammation associated with inhibiting JAK2/STAT3/Drp1-dependent mitochondrial fission in microglia. *Phytomedicine*, 59 (2019) 152922.
- Zhao T, Tang H, Xie L, Zheng Y, Ma Z, Sun Q & Li X, *Scutellaria baicalensis* Georgi. (Lamiaceae): a review of its traditional uses, botany, phytochemistry, pharmacology and toxicology. *J Pharm Pharmacol*, 71 (2019) 1353. <https://doi.org/10.1111/jphp.13129>.
- Liu J, Hou J, Jiang C, Li G, Lu H, Meng F & Shi L, Deep Sequencing of the *Scutellaria baicalensis* Georgi Transcriptome Reveals Flavonoid Biosynthetic Profiling and Organ-Specific Gene Expression. *PLoS One*, 10 (2015) e0136397.
- Tsai CL, Tsai CW, Chang WS, Lin JC & Hsia TC, Bau DT, Protective Effects of Baicalin on Arsenic Trioxide-induced Oxidative Damage and Apoptosis in Human Umbilical Vein Endothelial Cells. *In Vivo*, 35 (2021) 155.
- Wang M, Qiu S & Qin J, Baicalein induced apoptosis and autophagy of undifferentiated thyroid cancer cells by the ERK/PI3K/Akt pathway. *Am J Transl Res*, 11 (2019) 3341.
- Lian H, Hui Y, Xiaoping T, Wei T, Jiye X & Xiaolan Y, Baicalein suppresses the proliferation of human cervical cancer cells via Notch 1/Hes signaling pathway. *J Cancer Res Ther*, 15 (2019) 1216.
- Li K, Liang Y, Cheng A, Wang Q, Li Y, Wei H, Zhou C & Wan X, Antiviral Properties of Baicalin: a Concise Review. *Rev Bras Farmacogn*, 6 (2021) 1.
- Zhi H, Zhu H, Zhang Y, Lu Y, Li H & Chen DF, *In vivo* effect of quantified flavonoids-enriched extract of *Scutellaria baicalensis* root on acute lung injury induced by influenza A virus. *Phytomedicine*, 57 (2019) 105.
- Bai H, Yuan R, Zhang Z, Liu L, Wang X, Song X, Ma T, Tang J, Liu C & Gao L, Intra-articular Injection of Baicalein Inhibits Cartilage Catabolism and NLRP3 Inflammasome Signaling in a Posttraumatic OA Model. *Oxid Med Cell Longev*, 2 (2021) 6116890.
- Li N, Feng L, Tan Y, Xiang Y, Zhang R & Yang M, Preparation, Characterization, Pharmacokinetics and Biodistribution of Baicalin-Loaded Liposome on Cerebral Ischemia-Reperfusion after i.v. Administration in Rats. *Molecules*, 23 (2018) 1747.
- Fu Q, Gao L, Fu X, Meng Q & Lu Z, *Scutellaria baicalensis* Inhibits Coxsackievirus B3-Induced Myocarditis Via AKT and p38 Pathways. *J Microbiol Biotechnol*, 29 (2019) 1230.
- Kong Z, Shen Q, Jiang J, Deng M, Zhang Z & Wang G, Wogonin improves functional neuroprotection for acute cerebral ischemia in rats by promoting angiogenesis via TGF- $\beta$ 1. *Ann Transl Med*, 7 (2019) 639.
- Gong G, Wang H, Kong X, Duan R, Dong TTX & Tsim KWK, Flavonoids are identified from the extract of *Scutellariae Radix* to suppress inflammatory-induced angiogenic responses in cultured RAW 264.7 Macrophages. *Sci Rep*, 27 (2018) 17412.
- Zhao Y, Zhang L, Wu Y, Dai Q, Zhou Y, Li Z, Yang L Guo Q & Lu N, Selective anti-tumor activity of wogonin targeting the

- Warburg effect through stabilizing p53. *Pharmacol Res*, 135 (2018) 49.
- 21 Sharifi-Rad J, Herrera-Bravo J, Salazar LA, Shaheen S, Abdulmajid Ayatollahi S, Kobarfard F, Imran M, Imran A, Custódio L, Dolores López M, Schoebitz M, Martorell M, Kumar M, Ansar Rasul Suleria H & Cho WC, The Therapeutic Potential of Wogonin Observed in Preclinical Studies. *Evid Based Complement Alternat Med*, 15 (2021) 9935451.
  - 22 Ku SK & Bae JS, Antithrombotic activities of wogonin and wogonoside via inhibiting platelet aggregation. *Fitoterapia*, 98 (2014) 27.
  - 23 Lin CM, Chen YH, Ong JR, Ma HP, Shyu KG & Bai KJ, Functional role of wogonin in anti-angiogenesis. *Am J Chin Med*, 40 (2012) 415.
  - 24 Qu J, Zhang D, Liu F, Mao H, Ma Y, Yang Y, Li C, Qiu L, Geng X, Zhang J, Gao X, Chen L & Wang H, Vasodilatory Effect of Wogonin on the Rat Aorta and Its Mechanism Study. *Biol Pharm Bull*, 38 (2015) 1873.
  - 25 Chen G, Yang J, Lu G, Guo J & Dou Y, Limb remote ischemic post-conditioning reduces brain reperfusion injury by reversing eNOS uncoupling. *Indian J Exp Biol*, 52 (2014) 597.
  - 26 Maille N, Gokina N, Mandalà M, Colton I & Osol G, Mechanism of hydralazine-induced relaxation in resistance arteries during pregnancy: Hydralazine induces vasodilation via a prostacyclin pathway. *Vascul Pharmacol*, 78 (2016) 36.
  - 27 Mori A, Takeda K, Sakamoto K & Nakahara T, Activation of transient receptor potential vanilloid 4 channels dilates rat retinal arterioles through nitric oxide- and BKCa channel-dependent mechanisms in vivo. *Naunyn Schmiedeberg's Arch Pharmacol*, 393 (2020) 35.
  - 28 Villalba N, Sackheim AM, Nunez IA, Hill-Eubanks DC, Nelson MT, Wellman GC & Freeman K, Traumatic Brain Injury Causes Endothelial Dysfunction in the Systemic Microcirculation through Arginase-1-Dependent Uncoupling of Endothelial Nitric Oxide Synthase. *J Neurotrauma*, 34 (2017) 192.
  - 29 Buys ES, Zimmer DP, Chickering J, Graul R, Chien YT, Profy A, Hadcock JR, Masferrer JL & Milne GT, Discovery and development of next generation sGC stimulators with diverse multidimensional pharmacology and broad therapeutic potential. *Nitric Oxide*, 78 (2018) 72.
  - 30 Makhoul S, Walter E, Pagel O, Walter U, Sickmann A, Gambaryan S, Smolenski A, Zahedi RP & Jurk K, Effects of the NO/soluble guanylate cyclase/cGMP system on the functions of human platelets. *Nitric Oxide*, 76 (2018) 71.
  - 31 Chen C, Guo C, Gao J, Shi K, Cheng J, Zhang J, Chen S, Liu Y & Liu A, Vasorelaxant and antihypertensive effects of Tianshu Capsule on rats: An *in vitro* and *in vivo* approach. *Biomed Pharmacother*, 111 (2019) 188.
  - 32 García-Alonso D, Morgenstern-Kaplan D, Cohen-Welch A, Lozano-Cuenca J & López-Canales JS, Possible Mechanisms Involved in the Vasorelaxant Effect Produced by Anorexigenic Drugs in Rat Aortic Rings. *Med Sci (Basel)*, 27 (2019) 39.
  - 33 Li H, Shin SE, Seo MS, An JR, Ha KS, Han ET, Hong SH, Choi IW, Lee DS, Yim MJ, Lee JM, Jung ID, Firth AL, Han IY & Park WS, Inhibitory effect of the tricyclic antidepressant amitriptyline on voltage-dependent K<sup>+</sup> channels in rabbit coronary arterial smooth muscle cells. *Clin Exp Pharmacol Physiol*, 45 (2018) 205.
  - 34 Li H, Shin SE, Seo MS, An JR, Ha KS, Han ET, Hong SH, Kim J, Yim MJ, Lee JM, An TG, Jeon J, Lee SJ, Na SH & Park WS, Alterations of ATP-sensitive K<sup>+</sup> channels in human umbilical arterial smooth muscle during gestational diabetes mellitus. *Pflugers Arch*, 470 (2018) 1325.
  - 35 Zhu Y, Ye P, Chen SL & Zhang DM, Functional regulation of large conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels in vascular diseases. *Metabolism*, 83 (2018) 75.
  - 36 Dahal GR, Pradhan SJ & Bates EA, Inwardly rectifying potassium channels influence Drosophila wing morphogenesis by regulating Dpp release. *Development*, 144 (2017) 2771.
  - 37 Takahashi K, Hayashi S, Miyajima M, Omori M, Wang J, Kaihara K, Morimatsu M, Wang C, Chen J, Iribe G, Naruse K & Sokabe M, L-type calcium channel modulates mechanosensitivity of the cardiomyocyte cell line H9c2. *Cell Calcium*, 79 (2019) 68.
  - 38 Albarwani SA, Mansour F, Khan AA, Al-Lawati I, Al-Kaabi A, Al-Busaidi AM, Al-Hadhrani S, Al-Husseini I, Al-Siyabi S & Tanira MO, Aging Reduces L-Type Calcium Channel Current and the Vasodilatory Response of Small Mesenteric Arteries to Calcium Channel Blockers. *Front Physiol*, 7 (2016) 171.
  - 39 Duangjai A, Rukachaisirikul V, Sukpondma Y, Srimaroeng C & Muanprasat C, Antispasmodic Effect of Asperidine B, a Pyrrolidine Derivative, through Inhibition of L-Type Ca<sup>2+</sup> Channel in Rat Ileal Smooth Muscle. *Molecules*, 26 (2021) 5492.
  - 40 Jackson WF, Ion channels and the regulation of myogenic tone in peripheral arterioles. *Curr Top Membr*, 85 (2020) 19.
  - 41 Dietrich HH, Abendschein DR, Moon SH, Nayeb-Hashemi N, Mancuso DJ, Jenkins CM, Kaltenbronn KM, Blumer KJ, Turk J & Gross RW, Genetic ablation of calcium-independent phospholipase A(2)beta causes hypercontractility and markedly attenuates endothelium-dependent relaxation to acetylcholine. *Am J Physiol Heart Circ Physiol*, 298 (2010) H2208
  - 42 Shih HC & Yang L, Relaxant effect induced by wogonin from *Scutellaria baicalensis* on rat isolated uterine smooth muscle. *Pharm Biol*, 50 (2012) 760..