



Note

Royal jelly significantly alters inflammasome pathways in patients with chronic hepatitis B

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Royal jelly (RJ) plays immunomodulatory role in humans. Further, role played by inflammasomes against hepatitis B virus (HBV) and involvement in its complications are well known. Here, we evaluated the effects of RJ on the relative expression of apoptosis associated with speck-like protein (ASC), node like receptor (NLR) family pyrin domain containing 1 (NLRP1), NLRP3, S100 calcium binding protein A4 (S100A4), and S100A9, as the immune system-related molecules in patients with chronic hepatitis B infection. RJ was administered for 1 month (@1 g/day), to the patients with chronic hepatitis B infection. The relative expressions of ASC, NLRP1, NLRP3, S100A4 and S100A9 were evaluated using Real-Time PCR. The results showed that RJ increased the expression of ASC, but decreased the expression of NLRP1 in the patients with chronic hepatitis B infection. Relative expressions of NLRP3, S100A4, and S100A9 were not altered following treatment with RJ. There were no significant differences between men and women regarding the relative expression of the molecules. The results suggest that RJ can modulate immune responses via downregulation of NLRP1. The roles played by ASC in other pathways suggest that the upregulation of ASC could be associated with its immunomodulatory potential.

Keywords: *Apis mellifera*, Bee products, Gene expression.

Inflammasomes play key roles in the induction of immunity against viral infections, such as hepatitis B, and their related complications, including liver fibrosis¹. Among the inflammasomes, node like receptor (NLR) family pyrin domain containing 1 (NLRP1) and NLRP3 are the important intracellular sensors for the viral infections^{1,2}. It has been demonstrated that the inflammasomes participate in

neither defense against hepatitis B virus (HBV) nor induction of chronic inflammation, a main cause of liver cirrhosis and hepatocellular carcinoma (HCC)³. The most known members of inflammasomes, including NLRP1 and NLRP3, use apoptosis associated with speck-like protein (ASC) adaptor protein to activate their main target, caspase-1, as the protease for several proteins, including interleukin-1 beta (IL-1 β) and IL-18³. Previous investigations have shown the critical roles played by NLRP1 and NLRP3, and their adaptor protein, ASC against HBV and also its related liver complications⁴. Additionally, it has been demonstrated that internal damage associated molecular patterns (DAMPs) are the main ligands for innate immunity receptors, including inflammasomes⁵. S100 calcium binding protein A4 (S100A4) and S100A9 are the main members of DAMPs⁶. It has been reported that the S100 molecule expressions and functions had significant positive correlations with activation of NLRP1 and NLRP3⁷. Additionally, the roles played by S100A4 and S100A9 in the pathogenesis of chronic hepatitis B infections have been documented previously^{8,9}. Therefore, the factors that alter expression of the molecules may be considered as a therapeutic strategy against chronic hepatitis B infection.

Royal jelly (RJ), as a natural product from worker bees (*Apis mellifera*)¹⁰, is widely used for dietary supplements¹¹. RJ consists of several components, including water, protein, monosaccharides, fatty acids, 10-hydroxy-2-decenoic acid (10-HDA), antibacterial/antibiotic components and some vitamins¹². The component has potential antimicrobial, antioxidant and immunomodulatory roles¹³⁻¹⁶. RJ also promotes healthy aging and longevity¹², suppresses cancers¹⁷, epidermal stress¹⁸ and skeletal muscle atrophy¹⁹. Thus, it has been hypothesized that RJ may be useful to regulate immune responses in patients with chronic hepatitis B. Therefore, in this study, we evaluated the effect of royal jelly (RJ) treatment on the relative expressions of ASC, NLRP1, NLRP3, S100A4 and S100A9 in the CHB patients.

Material and Methods

Subjects

In this study, mRNA levels of ASC, NLRP1, NLRP3, S100A4, and S100A9 were analyzed in 30

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(13 men and 17 women) CHB patients, who were under RJ treatment. The patients were not under anti-HBV therapy and had normal liver enzyme serum levels and low HBV-DNA copy numbers. "Guide of Prevention and Treatment in Viral Hepatitis" was used to select the CHB diagnosis²⁰. To eliminate the interference factors, the patients with other microbial co-infection, receiving antiviral and immunosuppressive components, liver disorders, aged 18 to 55 years, pregnant or breastfeeding, and mental disorders were excluded from the project. The project protocol was approved by Kerman University of Medical Sciences Ethical Committee (IR.IAU. Kerman. REC.1400.010) and Iranian Registry of Clinical Trials (IRCT202106-20051640N1).

Blood samples were collected before and one month after RJ treatment in anticoagulant pre-treated tubes. RJ (Pars Asal Company, Shiraz, Iran) was administrated 1 g/day, based on our previous investigation²¹.

Evaluation of liver enzymes

The liver enzymes were evaluated using the commercial kits from MAN Company, Tehran, Iran, and based on the manufacturer's guidelines.

Extraction of mRNA

Total mRNA extraction was performed on the blood samples using a commercial kit (Karmania Pars Gene Company, Kerman, Iran). Briefly, the blood samples were lysed by the lysis buffer and then the precipitation solution was added to the lysate. After that, the component was added to the high absorbance column and after centrifuge, the column was washed using washing buffer. Finally, elution buffer was added to the column and total mRNA was separated from the column by centrifuge.

Complementary DNA (cDNA) synthesis

To convert the extracted mRNA to cDNA, a commercial kit from Karmania Pars Gene Company, Kerman, Iran, was used. Accordingly, 15 μ L master mix was mixed with extracted mRNA (1 μ g) and adjusted to 20 μ L by RNase/DNase free water. The component was incubated at 40°C for 3600 S and then 300 S at 70°C.

Relative expression of ASC, NLRP1, NLRP3, S100A4 and S100A9

Relative expression of ASC (CN# KPG-ASCRT), NLRP1 (CN# KPG-NLR1RT), NLRP3 (CN# KPG-NLR3RT), S100A4 (CN# KPG-S100A4RT), and S100A9 (CN# KPG-S100A9RT) was evaluated using the commercial kits from Karmania Pars Gene Company, Kerman, Iran.

Data analysis and statistical methods

Dependent paired 't' test under SPSS software (version 18) was used for data analysis, mRNA levels of ASC, NLRP1, NLRP3, S100A4 and S100A9 before and after therapy with RJ. *P* value was considered significant at <0.05.

Results

The results demonstrated that all the patients were positive for HBV-DNA and all of them were under 20/000 copy per mL. Liver enzymes were in normal ranges in all of the participants, before and after RJ treatment.

The treatment with RJ led to significant up and downregulation of ASC (*P* value= 0.024) and NLRP1 (*P* value= 0.050), respectively, in the CHB patients. Accordingly, relative expressions of ASC were increased after RJ treatment to 2.20-fold. Relative expressions of NLRP1 were decreased 7.83-fold after RJ treatment. The results demonstrated that relative expressions of NLRP3 (*P* value= 0.349), S100A4 (*P* value= 0.246) and S100A9 (*P* value= 0.280) before RJ treatment were not changed after RJ treatment. Fig. 1 shows the relative expression of ASC, NLRP1, NLRP3, S100A4 and S100A9 before and after RJ treatment.

The results demonstrated that there were no significant differences between men and women regarding mRNA levels of ASC, NLRP1, NLRP3,

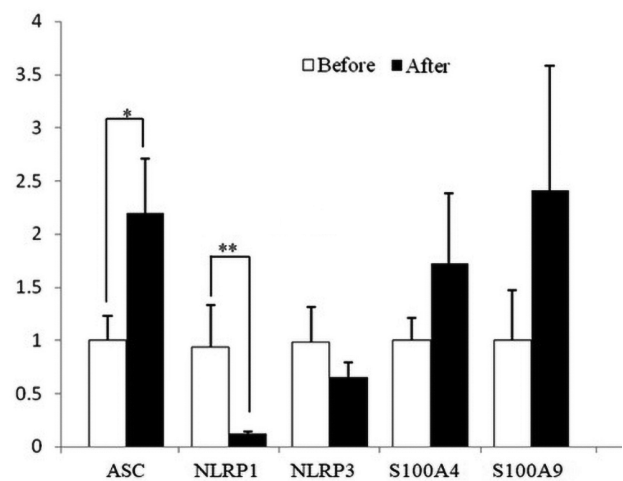


Fig 1 — Relative expression of apoptosis associated with speck-like protein (ASC), node like receptor (NLR) family pyrin domain containing 1 (NLRP1), NLRP3, S100 calcium binding protein A4 (S100A4), and S100A9 before and after treatment with royal jelly in the patients with chronic hepatitis B. [The figure shows that royal jelly therapy led to up regulation of ASC and down-regulation of NLRP1. * *P* value= 0.05, ***P* value= 0.025]

Table 1 — Relative expression of ASC, NLRP1, NLRP3, S100A4, and S100A9 in the men in comparison to women

Target gene	ASC B	ASC A	NLRP1 B	NLRP1 A	NLRP3 B	NLRP3 A	S100A4 B	S100A4 A	S100A9 B	S100A9 A
Men	0.89±0.38	1.39±0.50	0.86±0.57	0.12±0.03	0.51±0.36	0.31±0.11	1.70±0.97	3.88±2.32	1.56±0.75	10.28±8.98
Women	1.09±0.30	9.78±6.90	1.02±0.57	0.12±0.03	1.32±1.07	0.39±0.27	0.38±0.12	2.10±1.19	0.49±0.18	12.93±10.98
P value	0.687	0.273	0.850	0.987	0.506	0.801	0.226	0.479	0.209	0.866

[The table compares the relative expression of apoptosis associated with speck-like protein (ASC), node like receptor (NLR) family pyrin domain containing 1 (NLRP1), NLRP3, S100 calcium binding protein A4 (S100A4), and S100A9 in the men in comparison to women at both before (B) and after (A) RJ treatment. The t test revealed that relative expression of ASC, NLRP1, NLRP3, S100A4, and S100A9 were not different when compared men to women either before or after RJ treatment]

S100A4 and S100A9 either before or after RJ treatment. Table 1 illustrates the details of the relative expression of the molecules in the men and women.

Discussion

The results demonstrate that receiving RJ for a period of one month was associated with increased expression of ASC and decreased expression of NLRP1. Previous investigations proved the critical roles played by NLRP1 in the pathogenesis of chronic hepatitis B⁴. Our results show that RJ can modulate expression of NLRP1 and it may be hypothesized that the natural component may confer immunoregulatory effects in the patients with chronic hepatitis B. It has been reported that ASC not only makes a link between the inflammasomes and caspase-1, but also it plays other roles in immune system, including regulation of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) pathway and activation of mitogen-activated protein kinase (MAPK), as well as regulation of the dedicator of cytokinesis 2 (Dock2) mRNA stability²². MAPK, and Dock2 pathways play significant roles during immune responses and make immune response more effective against microbes, including viruses. However, it has been reported that RJ can suppress phosphorylation of NF-κB and other related signaling pathways, such as p38 and c-Jun NH2-terminal protein kinase (JNK)²³⁻²⁶. Hence, it may be hypothesized that although mRNA levels of ASC were increased following RJ treatment, it may not result in activation of the pathway. However, based on the immunomodulatory effects of RJ, increased expression of ASC may show the immunoregulatory effects of the molecules, except its known pro-inflammatory roles. Thus, it seems that RJ via down-regulation of NLRP1 can modulate the pathogenesis of the molecule during chronic form of hepatitis B and it may be associated with increased functions of other pathways in ASC dependent ones.

Based on our knowledge, this is the first study on RJ treatment in the chronic hepatitis B and on the

expression of ASC, NLRP1, NLRP3, S100A4, and S100A9 molecules. Our results revealed that RJ was unable to change expression of NLRP3 as the most known inflammasome, and also S100A4 and S100A9, as DAMPs. However, the increased sample size may be associated with alteration in the expression of these molecules. Further investigation with higher sample size and increased RJ doses may clarify the mechanism associated with alteration in expression of the molecules. In parallel with our results, investigations revealed that RJ can relieve the necrotic hepatocytes via downregulation of tumor necrosis factor (TNF) -α, mixed lineage kinase domain-like protein (MLKL) and intracellular reactive species, the most important agents of hepatic necrosis, and suppresses HBV entry and replication in the hepatocytes^{27,28}.

The results also demonstrated that relative expression of ASC, NLRP1, NLRP3, S100A4, and S100A9 were not different in the men and women. Our previous investigations also proved the fact that gender did not affect expression of immune related molecules in the patients with chronic hepatitis B²⁹⁻³¹. Therefore, it appears that gender cannot be considered as an important factor to alter expression of the immune system-related molecules in the Iranian chronic hepatitis B patients.

Conclusion

The above study has demonstrated that the Royal Jelly (RJ) from the worker bees of *Apis mellifera* could modulate immune responses in patients suffering from chronic hepatitis B through downregulation of NLRP1. Due to the roles played by ASC in other pathways, upregulation of the ASC might be associated with immunomodulatory roles. It has also been shown that the gender is not a potential factor for relative expression of ASC, NLRP1, NLRP3, S100A4 and S100A9.

Conflict of Interest

Author declares no competing interests.

References

- 1 Li Z & Jiang J, The NLRP3 inflammasome mediates liver failure by activating procaspase-1 and pro-IL-1 β and regulating downstream CD40-CD40L signaling. *J Int Med Res*, 49 (2021) 3000605211036845. doi: 10.1177/03000605211036845.
- 2 Askari A, Nosratabadi R, Khaleghinia M, Zainodini N, Kennedy D, Shabani Z & Arababadi MK, Evaluation of NLR4, NLRP1, and NLRP3, as components of inflammasomes, in chronic hepatitis B virus-infected patients. *Viral Immunol*, 29 (2016) 496. doi: 10.1089/vim.2016.0045.
- 3 Dadmanesh M, Ranjbar MM & Ghorban K, Inflammasomes and their roles in the pathogenesis of viral hepatitis and their related complications: An updated systematic review. *Immunol Lett*, 208 (2019) 11. doi: 10.1016/j.imlet.2019.03.001.
- 4 Mameros AG, Role of inflammasome activation in neovascular age-related macular degeneration. *FEBS J*, 12 (2021) doi: 10.1111/febs.16278. doi: 10.1111/febs.16278.
- 5 Choltus H, Lavergne M, De Sousa Do Outeiro C, Coste K, Belleville C, Blanchon L & Sapin V, Pathophysiological implication of pattern recognition receptors in fetal membranes rupture: RAGE and NLRP inflammasome. *Biomedicines*, 9 (2021) 1123. doi: 10.3390/biomedicines9091123.
- 6 Imbalzano E, Mandraffino G, Casciaro M, Quartuccio S, Saitta A & Gangemi S, Pathophysiological mechanism and therapeutic role of S100 proteins in cardiac failure: a systematic review. *Heart Fail Rev*, 21 (2016) 463. doi: 10.1007/s10741-016-9529-8.
- 7 Zhang J, Hou S, Gu J, Tian T, Yuan Q, Jia J, Qin Z & Chen Z, S100A4 promotes colon inflammation and colitis-associated colon tumorigenesis. *Oncotarget*, 7 (2018) 1461301. doi: 10.1080/2162402X.2018.1461301.
- 8 Zhu K, Huang W, Wang W, Liao L, Li S, Yang S, Xu J, Li L, Meng M, Xie Y, He S, Tang W, Zhou H, Liang L, Gao H, Zhao Y, Hou Z, Tan J & Li R, Up-regulation of S100A4 expression by HBx protein promotes proliferation of hepatocellular carcinoma cells and its correlation with clinical survival. *Gene*, 749 (2020) 144679. doi: 10.1016/j.gene.2020.144679.
- 9 Wu R, Zhang Y, Xiang Y, Tang Y, Cui F, Cao J, Zhou L, You Y & Duan L, Association between serum S100A9 levels and liver necroinflammation in chronic hepatitis B. *J Transl Med*, 16 (2018) 83. doi: 10.1186/s12967-018-1462-2.
- 10 Kamakura M, Royalactin induces queen differentiation in honeybees. *Nature*, 473 (2011) 478. doi: 10.1038/nature10093.
- 11 Ramadan MF & Al-Ghamdi A, Bioactive compounds and health-promoting properties of royal jelly: A review. *J Funct Food*, 4 (2012) 39. doi:10.1016/j.jff.2011.12.007.
- 12 Kunugi H & Ali AM, Royal Jelly and Its Components Promote Healthy Aging and Longevity: From Animal Models to Humans. *Int J Mol Sci*, 20 (2019) 4662. doi: 10.3390/ijms20194662.
- 13 Pavel CI, Marghitas LA, Bobis O, Dezmirean DS, Sapcaliu A, Radoi I & Madas MN, Biological activities of royal jelly-review. *Sci Papers Anim Sci Biotech*, 44 (2011) 108.
- 14 El-Gayar MH, Aboshanab KM, Aboulwafa MM & Hassouna NA, Antivirulence and wound healing effects of royal jelly and garlic extract for the control of MRSA skin infections. *Wound Med*, 13 (2016) 18. doi.org/10.1016/j.wndm.2016.05.004.
- 15 Nascimento AP, Moraes LA, Ferreira NU, Moreno Gde P, Uahib FG, Barizon EA & Berretta AA, The lyophilization process maintains the chemical and biological characteristics of Royal jelly. *Evid Based Complement Alternat Med*, 2015 (2015) 825068. doi.org/10.1155/2015/825068.
- 16 Delkhoshe-Kasmaie F, Malekinejad H, Khoramjouy M, Rezaei-Golmishah A & Janbaze-Acyabar H, Royal jelly protects from taxol-induced testicular damages via improvement of antioxidant status and up-regulation of E2f1. *Syst Biol Reprod Med*, 60 (2014) 80. doi: 10.3109/19396368.2013.852271.
- 17 Miyata Y & Sakai H, Anti-Cancer and Protective Effects of Royal Jelly for Therapy-Induced Toxicities in Malignancies. *Int J Mol Sci*, 19 (2018) 3270. doi: 10.3390/ijms19103270.
- 18 Okumura N, Ito T, Degawa T, Moriyama M & Moriyama H, Royal Jelly Protects against Epidermal Stress through Upregulation of the NQO1 Expression. *Int J Mol Sci*, 22 (2021) 12973. doi: 10.3390/ijms222312973.
- 19 Shirakawa T, Miyawaki A, Matsubara T, Okumura N, Okamoto H, Nakai N, Rojasawasthien T, Morikawa K, Inoue A, Goto A, Washio A, Tsujisawa T, Kawamoto T & Kokabus, Daily Oral Administration of Protease-Treated Royal Jelly Protects Against Denervation-Induced Skeletal Muscle Atrophy. *Nutrients*, 12 (2020) 3089. doi: 10.3390/nu12103089.
- 20 Liu HG, Chen WW, Fan ZP, Yang HY, Shi M, Zhang Z, Luan SS, Zhang H, Lu P, Tien P & Wang FS, The high prevalence of the I27 mutant HBeAg18-27 epitope in Chinese HBV-infected patients and its cross-reactivity with the V27 prototype epitope. *Clin Immunol*, 125 (2007) 337. doi: 10.1016/j.clim.2007.06.010.
- 21 Pourmobini H, Kazemi Arababadi M, Salahshoor MR, Roshankhah S, Taghavi MM, Taghipour Z & Shabanizadeh A, The effect of royal jelly and silver nanoparticles on liver and kidney inflammation. *Avicenna J Phytomed*, 11 (2021) 218.
- 22 Hassan H & Amer AO, Cell intrinsic roles of apoptosis-associated speck-like protein in regulating innate and adaptive immune responses. *ScientificWorldJournal*, 11 (2011) 2418. doi: 10.1100/2011/429192.
- 23 Ali AM & Kunugi H, Bee honey protects astrocytes against oxidative stress: A preliminary in vitro investigation. *Neuropharmacol Rep*, 39 (2019) 312. doi: 10.1002/npr.212079.
- 24 Kawahata I, Xu H, Takahashi M, Murata K, Han Y, Yamaguchi Y, Fujii A, Yamaguchi K & Yamakuni T, Royal jelly coordinately enhances hippocampal neuronal expression of somatostatin and neprilysin genes conferring neuronal protection against toxic soluble. *J Function Food*, 51 (2018) 28. doi.org/10.1016/j.jff.2018.10.006.
- 25 Mohamed AA, Galal AA & Elewa YH, Comparative protective effects of royal jelly and cod liver oil against neurotoxic impact of tartrazine on male rat pups brain. *Acta Histochem*, 117 (2015) 649. doi: 10.1016/j.acthis.2015.07.002.
- 26 You MM, Chen YF, Pan YM, Liu YC, Tu J, Wang K & Hu FL, Royal jelly attenuates LPS-induced inflammation in BV-2 microglial cells through modulating NF- κ B and p38/JNK signaling pathways. *Mediators Inflamm*, 2018 (2018) 7834381. doi: 10.1155/2018/7834381.
- 27 Abu-Serie MM & Habashy NH, Two purified proteins from royal jelly with in vitro dual anti-hepatic damage potency:

- Major royal jelly protein 2 and its novel isoform X1, *Int J Biol Macromol*, 128 (2019) 782. doi: 10.1016/j.ijbiomac.2019.01.210.
- 28 Marwa M & Abu-Serie NH, Major royal-jelly protein 2 and its isoform X1 are two novel safe inhibitors for hepatitis C and B viral entry and replication. *Int J Biol Macromol*, 141 (2019) 1087. doi: 10.1016/j.ijbiomac.2019.09.080.
- 29 Bahramabadi R, Fathollahi MS, Hashemi SM, Arababadi AS, Arababadi MS, Yousefi-Daredor H, Bidaki R, Khaleghinia M, Bakhshi MH, Yousefpoor Y, Torbaghan YE & Arababadi MK, Serum levels of IL-6, IL-8, TNF- α , and TGF- β in chronic HBV-infected patients: Effect of depression and anxiety. *Lab Med*, 49 (2017) 41. doi: 10.1093/labmed/lmx064.
- 30 Safari-Arababadi M, Modarressi MH & Arababadi MK, Up-regulation of RIP1 and IPS-1 in chronic HBV infected patients. *Genet Mol Biol*, 42 (2019) 337. doi: 10.1590/1678-4685-GMB-2018-0071.
- 31 Ebrahim M, Mirzaei V, Bidaki R, Shabani Z, Daneshvar H, Karimi-Googheri M, Khaleghinia M, Afroz MR, Yousefpoor Y & Arababadi MK, Are RIG-1 and MDA5 expressions associated with chronic HBV infection? *Viral Immunol*, 28 (2015) 504. doi: 10.1089/vim.2015.0056.